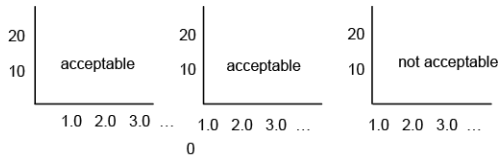


Mark scheme: Biological Molecules - PAG's

Question	Answer/Indicative content	Marks	Guidance
1	C ✓	1	
	Total	1	
2	C	1	
	Total	1	
3	D	1	
	Total	1	
4	A	1	
	Total	1	
5	<p><i>two from</i> add biuret / NaOH and CuSO₄, solution / reagent to urine (1) observe colour change (from blue to purple) (1) compare with, control / blank (urine containing no protein) (1)</p>	2	IGNORE biuret test unqualified.
	Total	2	
6	<p><i>idea that</i> more than one leaf should be tested ✓</p> <p>in case the leaf used was atypical ✓</p> <p>sucrose is a non-reducing sugar ✓</p> <p>if method is intended to measure sucrose then boiling with HCl is necessary ✓</p> <p>other sugars / glucose, also present in leaf ✓</p> <p>chlorophyll / other pigments, in leaf ✓</p> <p>green / colour, of pigments will interfere with colorimetry results ✓</p> <p>blue filter is the wrong type of filter / should have used red filter ✓</p>	4 max	
	Total	4	


7	a	i	<p>1 appropriate scale chosen</p> <p>and x axis labelled glucose concentration (mmol dm^{-3})</p> <p>and y axis labelled mean % absorbance ✓</p> <p>2 points plotted correctly ✓</p> <p>3 straight line of best fit drawn on graph (not extending beyond the plot points) ✓</p>	<p>1 IGNORE presence or absence of 0 at origin(s) unless either axis is deemed to have started above 0</p>  <p>2</p> <table border="1" data-bbox="917 544 1370 685"> <tr> <td>x axis glucose concentration (mmol dm^{-3})</td> <td>1.0</td> <td>2.0</td> <td>3.0</td> <td>4.0</td> <td>5.0</td> <td>6.0</td> </tr> <tr> <td>y axis mean % absorbance</td> <td>67</td> <td>54</td> <td>47</td> <td>41</td> <td>26</td> <td>16</td> </tr> </table> <p>Centre of cross or dot within + or – half a small square</p> <p>Centre of cross or dot within + or – half a small square one error in the plotted points ALLOW one error in the plotted points Points for glucose concs 1, 3, 5 & 6 mmol dm^{-3} should be in a straight line.</p> <p>Points for glucose concs 2, 3 & 4 mmol dm^{-3} should be in a straight line with a shallower gradient</p> <p>Note: A bar chart will only be able to access mp 2</p> <p>Examiner's Comments Few candidates scored all three marks. Those who failed to quote the y axis variable correctly as 'mean % absorbance', chose a scale that was too small for the available space or who used a non-linear scale were not awarded a mark. Point plotting was usually accurate and this mark was frequently awarded. As the data showed a calibration curve a line of best fit should have been drawn. Many extrapolated their line of best fit inappropriately. The following guidance can be found in the Mathematical Skills Handbook: 'Where the purpose of the graph is to present the data within the experimental range only (with the possibility for deriving predicted values through interpolation) no extrapolation is needed and the line should be confined to the range of the independent variable.'</p>	x axis glucose concentration (mmol dm^{-3})	1.0	2.0	3.0	4.0	5.0	6.0	y axis mean % absorbance	67	54	47	41	26	16
x axis glucose concentration (mmol dm^{-3})	1.0	2.0	3.0	4.0	5.0	6.0												
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		<p>ii</p> <p>find the absorbance (of the juice using the colorimeter) ✓</p> <p>(from the graph) find the concentration that corresponds to this absorbance ✓</p> <p>follow the, absorbance value / value on y axis, across to, line of best fit / (calibration) curve, and then down to the, concentration / x axis ✓</p>	<p>2 max</p>	<p>ACCEPT vertical and horizontal for x and y</p> <p>Examiner's Comments Candidates may have been unfamiliar with the use of calibration curves or else struggled to describe their use in a logical, step-wise answer, as the quality of answers was often poor. Few candidates integrated the information across all of the question and realised that they would have to find the absorbance values of A, B and C using the colorimeter before locating these values on the y axis, ruling a line across to the line of best fit and reading down to the x axis value for concentration. Some scored the mark for the general idea that the curve could be used to find the concentration that corresponds to a particular absorbance. Weaker answers described how the glucose concentration would affect the absorbance without attempting to answer the question.</p>
	<p>b</p> <p>i</p>	<ol style="list-style-type: none"> taste the fruit juices to see how sweet they are ✓ place a sample of each fruit juice in a biosensor and take the reading or test each fruit juice with, Benedict's / diastix / clinistix / (diagnostic) test strip and observe colour(s) ✓ obtain rank order for, sweetness / fruit juice glucose concentration 	<p>4 max</p>	<ol style="list-style-type: none"> could be in the context of different juices or a series of dilutions of the same juice (to give different glucose concentrations) or a series of glucose concentrations ACCEPT semi-quantitative test for reducing sugar Benedict's tests on each fruit juice and weigh mass of precipitate formed for each juice perform ACCEPT plausible way of determining glucose concentration e.g. relative density / specific gravity / mass change as a result of osmosis Benedict's – blue to red with increasing concentration diastix – green / blue to red clinistix - green / blue to red or pink to (dark) purple

		<p>4. compare rank orders (of fruit juices) for sweetness and glucose concentration ✓</p> <p>5. how a variable was controlled during, taste / glucose concentration, test ✓</p>	<p>4. ACCEPT the use of a statistical test if rank orders for both are numerical</p> <p>5. e.g. use same, number of drops / volumes, of fruit juice cleanse palate between juices blind taste test / stated way to avoid bias tasted by a number of subjects (and results pooled) keep test strip in sample for same length of time add excess Benedict's heat for same length of time / at the same temperature (Benedict's only) filter precipitate in same way (semi-quantitative Benedict's only)</p> <p>Examiner's Comments Most candidates showed good knowledge of the Benedict's test though there was some confusion over whether it tests for a reducing or non- reducing sugar. A mark was often scored for controlling a variable during the test such as the volume of juice or Benedict's solution, or the heating time or temperature. Fewer candidates went on to describe a taste test, either of different juices (A, B and C), or of dilutions of a juice or of a range of glucose concentrations. Answers that discussed taste tests tended to do well and often achieved both the method mark and the idea of ranking the results to give an order of sweetness. Some discussed tasting the juices but did not state 'to see how sweet they were'. The most insightful answers compared the rank order of sweetness with the rank order found with the Benedict's test.</p>
	ii	<p>tasting is, subjective / (only) qualitative / not quantitative or hard to quantify sweetness or people may have different, judgement / opinion / taste buds ✓</p> <p>colour judgement (in Benedict's) is subjective ✓</p>	<p>IGNORE accuracy / reliability</p> <p>1 max ACCEPT ref to biased opinion</p>

		(juice) may contain, sucrose / fructose / other (named) sugar / (artificial) sweetener ✓		<p>ACCEPT sensible ref to acidity in juice masking sweetness</p> <p>IGNORE ref to 'other ingredients' unqualified</p> <p>Examiner's Comments Where candidates' train of thought had led them to discussing a taste test in the previous answer they almost always came up with a reason why the results might not support the hypothesis, due to different (non-glucose) sugars being present in the juice or other sweeteners or fruit acids affecting the perception of sweetness of the juice. The subjective nature of the taste test or of judging the final colour of the Benedict's test also scored a mark for many.</p>
		Total	10	
8		<p>1 <u>zero</u> the colorimeter / set to <u>zero</u> }</p> <p>2 using <u>blank</u> }</p> <p>3 use red filter }</p> <p>4 use known concentrations (of lactose) }</p> <p>5 (produce) serial / series, dilutions }</p> <p>6 construct calibration curve }</p> <p>7 test unknown sample (using the same method) }</p> <p>8 use / read from, graph / calibration curve, to determine (unknown) concentration }</p>	4 max	<p>ALLOW calibrate to zero</p> <p>3 ALLOW red light / orange filter</p> <p>4 ALLOW a list of stated concentrations</p> <p>5 ALLOW clear description</p> <p>6 ALLOW plot concentration against, transmission / absorbance</p> <p>8 Cannot be assumed from mp 6</p> <p>Examiner's Comments This question differentiated well between candidates. Candidates should be familiar with this type of practical from the practical endorsement (PAG 5), and most ought to have carried out a similar practical activity.</p> <p>A large proportion of candidates began their answers with detailed descriptions of various aspects of the Benedict's test that were not relevant to the calibration process but then went on to score marks with relevant descriptions of the calibration procedure. Some focused on describing a Benedict's test or explaining the principle which they were not asked to do, and so did not receive much, if any, credit. All of the marking points were seen but serial dilutions were less commonly</p>

				<p>suggested despite these being a feature of OCR's PAG activities. It is worth noting that in order to construct a calibration curve, more than one known concentration needs to be used. Centres are reminded that the practical components of the syllabus are integral to students being able to apply their theoretical learning; performing these practical activities will enable candidates to relate to these elements when tested in the examinations.</p>
		Total	4	
9		<p>Phloem = B AND contains sucrose / non-reducing sugar ✓ non-reducing sugar / sucrose, hydrolysed / broken down, to monosaccharides ✓</p> <p>Liver = A AND does not contain starch / gives negative result for iodine test ✓</p>	3	<p>ALLOW non-reducing sugars broken down to, reducing sugars / named monosaccharide</p> <p>ALLOW 'colour after iodine added was yellow'</p> <p><u>Examiner's Comments</u></p> <p>Many candidates identified tissue B as phloem since it contained sucrose, or a non-reducing sugar, which would result in a red precipitate with Benedict's reagent after treatment with hydrochloric acid. Fewer mentioned that this treatment would hydrolyse sucrose into its monosaccharide constituents. Some candidates lost the mark for stating that sucrose is a reducing sugar.</p> <p>Most candidates correctly identified tissue A as liver due to the fact that it contained no starch resulting in a negative result for the iodine test. Lower ability candidates identified C as the liver stating that it would contain both glycogen and reducing sugars such as glucose, but ignoring the fact that tissue C also contained starch.</p>
		Total	3	
10	i	<p>non-reducing , sugars / disaccharides ✓</p>	<p>1 (AO3.2)</p>	<p>ALLOW sucrose / cellulose / vitamins IGNORE minerals / ions / fibre</p> <p><u>Examiner's Comments</u></p>

		<p>The majority of candidates gave 'non-reducing sugar' as the correct response for this part of the question with a few opting for 'vitamins' or the correctly named disaccharide, 'sucrose', which were also credited.</p>																		
	<p>ii</p> <table border="1" style="margin-left: auto; margin-right: auto;"> <thead> <tr> <th data-bbox="331 728 389 801">Test tube</th> <th data-bbox="389 728 619 801">Observations</th> <th data-bbox="619 728 756 801">Conclusion</th> </tr> </thead> <tbody> <tr> <td data-bbox="331 801 389 891">1</td> <td data-bbox="389 801 619 891">(pale) purple / lilac / violet / mauve</td> <td data-bbox="619 801 756 891">Protein present</td> </tr> <tr> <td data-bbox="331 891 389 1010">2</td> <td data-bbox="389 891 619 1010">Yellow colour</td> <td data-bbox="619 891 756 1010">reducing sugar (present)</td> </tr> <tr> <td data-bbox="331 1010 389 1128">3</td> <td data-bbox="389 1010 619 1128">Pale brown colour</td> <td data-bbox="619 1010 756 1128">no / very little , starch (present)</td> </tr> <tr> <td data-bbox="331 1128 389 1317">4</td> <td data-bbox="389 1128 619 1317">(turns) white / cloudy / milky OR (forms white) suspension / emulsion</td> <td data-bbox="619 1128 756 1317">Fat present</td> </tr> <tr> <td data-bbox="331 1317 389 1471">5</td> <td data-bbox="389 1317 619 1471">pink</td> <td data-bbox="619 1317 756 1471">Glucose concentration small (15 mg dm⁻³)</td> </tr> </tbody> </table> <p style="text-align: center; margin-left: 150px;">✓</p> <p style="text-align: center; margin-left: 180px;">✓</p>	Test tube	Observations	Conclusion	1	(pale) purple / lilac / violet / mauve	Protein present	2	Yellow colour	reducing sugar (present)	3	Pale brown colour	no / very little , starch (present)	4	(turns) white / cloudy / milky OR (forms white) suspension / emulsion	Fat present	5	pink	Glucose concentration small (15 mg dm ⁻³)	<p>1 mark per correct column</p> <p>IGNORE monosaccharides</p> <p>DO NOT ALLOW precipitate</p> <p>IGNORE any qualifications / shades of colour</p> <p><u>Examiner's Comments</u></p> <p>There were some excellent responses from candidates who were able to complete both 'observation' and 'conclusion' columns demonstrating good knowledge of tests for biological 'food' molecules. However, some candidates did confuse the different tests with a common error to give brick-red precipitate for the expected observation in Tube 1.</p> <p style="text-align: center;">2 (AO3.2)</p> <p style="text-align: center;"> OCR support</p> <p>Practical work should be an integral part of the study of Biology. The practicals provided by OCR to support the practical endorsement include Practical Activity Group (PAG) 9 in which there are a series of qualitative tests. PAG activities are available on OCR interchange: https://interchange.ocr.org.uk/Downloads/PAG9.zip</p>
Test tube	Observations	Conclusion																		
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	<p>iii</p> <p>(result using colorimeter will be) <u>quantitative</u> OR not subjective / less affected by human error / no bias ✓</p>	<p style="text-align: center;">1 (AO3.4)</p> <p>IGNORE accurate / valid ALLOW is objective</p>																		

				<p>Examiner's Comments</p> <p>The majority of candidates understood the use of the colorimeter in giving quantitative or non-subjective results. However, there are still many candidates using the terms accuracy and precision in the wrong context (such as here) when responding to AO3 practical-based questions.</p> <p> OCR support</p> <p>Appendix 4 of the Practical Skills Handbook, provides information on terms used in measurement and conventions for recording and processing experimental measurements. This is in line with the 'The Language of measurement' booklet: https://www.ocr.org.uk/Images/294468-biology-practical-skills-handbook.pdf</p>												
		Total	4													
1 1		<table border="1"> <thead> <tr> <th>Improvement</th> <th>Justification</th> </tr> </thead> <tbody> <tr> <td>Use a colorimeter with a digital display showing absorbance units to 3 decimal places.</td> <td>To assess repeatability</td> </tr> <tr> <td>Check the zero value of the colorimeter with purified water before use.</td> <td>To assess reproducibility</td> </tr> <tr> <td>For each concentration, repeat the measurement of the rate of reaction three times and calculate a mean.</td> <td>To reduce systematic error</td> </tr> <tr> <td>Ask students in several schools to carry out the same investigation.</td> <td>To reduce random error (uncertainty)</td> </tr> <tr> <td></td> <td>To increase resolution</td> </tr> </tbody> </table>	Improvement	Justification	Use a colorimeter with a digital display showing absorbance units to 3 decimal places.	To assess repeatability	Check the zero value of the colorimeter with purified water before use.	To assess reproducibility	For each concentration, repeat the measurement of the rate of reaction three times and calculate a mean.	To reduce systematic error	Ask students in several schools to carry out the same investigation.	To reduce random error (uncertainty)		To increase resolution	4 (AO2.3)	<p>One mark per correct line DO NOT ALLOW more than one line per box</p>
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		Total	4													
1 2		<p>(carry out) Benedict's test / described ✓</p> <p>(if test for reducing sugar negative) boil with (dilute) HCl and (re)test (with Benedict's) ✓</p>	2 (AO2.7)	<p>ALLOW 'add Benedict's (solution)'</p> <p>Examiner's Comments</p> <p>Most candidates answered (b)(ii) and (c) well, showing a good understanding of the Krebs cycle and chemiosmosis. Among some</p>												

			<p>candidates, there was some confusion of how the proton gradient is established and few candidates mentioned that energy provided by the flow of protons through ATP synthase is used to join ADP and Pi to form ATP.</p> <p>Very few candidates scored 2 marks for (a). This question required a candidate to describe performing the Benedict's test for a reducing sugar which would produce a negative result and then repeating the test after boiling with Hydrochloric acid. Boiling with Hydrochloric acid before performing the first Benedict's test would not enable you to differentiate a positive test for a reducing sugar from a positive test for a non – reducing sugar.</p> <p>Very few candidates understood the meaning of the RQ values in Fig 4.3. Most candidates linked the RQ value to the amount of respiration taking place or the relative amounts of carbon dioxide being released compared to oxygen consumed. Few were able to link the RQ value to the type of respiratory substrate mainly being used at 0W and 50W, although many realised the high RQ value at 250W indicated that anaerobic respiration was taking place.</p> <div data-bbox="895 1279 975 1357" style="text-align: center;"> </div> <p style="text-align: center;">AfL</p> <p>As well as calculating RQ values, encourage candidates to interpret RQ values in terms of types of respiratory substrate being used and whether respiration is aerobic or anaerobic.</p>
		Total	2